

## Aporphine Glycosides from *Stephania cepharantha* Seeds

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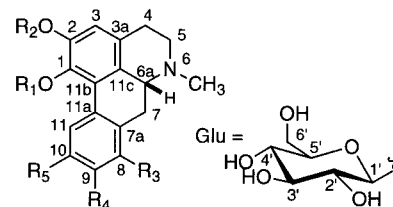
Two new aporphine glycosides, stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**) and *N*-methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**), were isolated from the seeds of *Stephania cepharantha* cultivated in Japan, together with 16 known alkaloids. The structures of **1** and **2** were spectroscopically determined by comparison of their  $^1\text{H}$  and  $^{13}\text{C}$  NMR data with those of stesakine (**11**) and *N*-methylasimilobine (**12**), respectively.

The alkaloid constituents from the root tubers,<sup>1</sup> leaves,<sup>2</sup> and stems<sup>3</sup> of *Stephania cepharantha* Hayata (Menispermaceae) cultivated in Japan were reported in previous papers. The alkaloid constituents of the seeds have been investigated by Kunitomo et al., who isolated eight alkaloids: six aporphines and two bisbenzylisoquinolines.<sup>4</sup> In our investigation, two new aporphine glycosides, stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**) and *N*-methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**), were obtained, together with 16 known alkaloids.<sup>5</sup> This paper describes the isolation and structure elucidation of **1** and **2**.

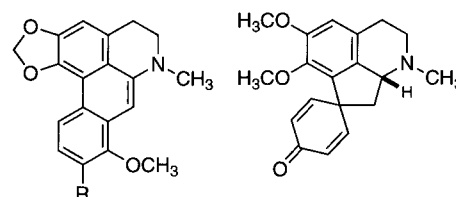
### Results and Discussion

Seeds of *S. cepharantha* were successively extracted with *n*-hexane and MeOH. The *n*-hexane extract was repeatedly subjected to a combination of column chromatography, preparative TLC, and crystallization, to afford 10 known aporphine alkaloids: dehydrostephanine (**3**),<sup>4</sup> dehydrocrebanine (**4**),<sup>6</sup> stephanine (**5**),<sup>7</sup> crebanine (**6**),<sup>7</sup> (-)-dicentrine (**7**),<sup>8</sup> (-)-roemerine (**8**),<sup>9</sup> (-)-isolaureline (**9**),<sup>10</sup> nuciferine (**10**),<sup>11</sup> stesakine (**11**),<sup>6</sup> and *N*-methylasimilobine (**12**).<sup>11</sup> The MeOH extract was fractionated, and the alkaloid-containing fraction was purified using the same methods as for the fractionation of the *n*-hexane extract, to give stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**) and *N*-methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**), as well as nine known alkaloids: four aporphines, **6**, **11**, **12**, and (-)-cassythicine (**13**);<sup>12</sup> one proaporphine, pronuciferine (**14**);<sup>13</sup> and four bisbenzylisoquinolines, cepharanthine,<sup>14</sup> cepharanoline,<sup>15</sup> isotetrandrine,<sup>16</sup> and berbamine.<sup>17</sup>

Glycoside **1** was obtained as an amorphous powder. The molecular formula of  $\text{C}_{25}\text{H}_{29}\text{NO}_9$  was deduced by FABMS. The EIMS showed a molecular ion peak at  $m/z$  487 and strong ion peaks at  $m/z$  325  $[\text{M} - 162]^+$  and 324  $[\text{M} - 163]^+$ , indicating that **1** was probably an alkaloid glycoside.<sup>18,19</sup> The  $^1\text{H}$  NMR spectrum showed signals of one *N*-methyl group ( $\delta$  2.48), one methoxy group ( $\delta$  4.01), methylenedioxy protons ( $\delta$  5.98, 6.11), one singlet aromatic proton ( $\delta$  6.60), and a set of coupled aromatic protons ( $\delta$  7.71, 8.01); the spectrum was similar to that of stesakine (**11**), except for signals assigned to the sugar moiety. The  $^{13}\text{C}$  NMR spectrum showed 25 signals, of which five methines and one methylene were assigned to the  $\beta$ -glucopyranose moiety; the other 19 signals were similar to those of **11**. These data suggested that **1** was an aporphine glycoside, with **11** as the aglycon and  $\beta$ -glucopyranose as the sugar



	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	R <sub>4</sub>	R <sub>5</sub>
<b>1</b>	-CH <sub>2</sub> -		OCH <sub>3</sub>	OGlu	H
<b>2</b>	CH <sub>3</sub>	Glu	H	H	H
<b>5</b>	-CH <sub>2</sub> -		OCH <sub>3</sub>	H	H
<b>6</b>	-CH <sub>2</sub> -		OCH <sub>3</sub>	OCH <sub>3</sub>	H
<b>7</b>	-CH <sub>2</sub> -		H	OCH <sub>3</sub>	OCH <sub>3</sub>
<b>8</b>	-CH <sub>2</sub> -		H	H	H
<b>9</b>	-CH <sub>2</sub> -		H	OCH <sub>3</sub>	H
<b>10</b>	CH <sub>3</sub>	CH <sub>3</sub>	H	H	H
<b>11</b>	-CH <sub>2</sub> -		OCH <sub>3</sub>	OH	H
<b>12</b>	CH <sub>3</sub>	H	H	H	H
<b>13</b>	-CH <sub>2</sub> -		H	OH	OCH <sub>3</sub>



**3** R = H, **4** R = OCH<sub>3</sub>

**14**

moiety. The sugar linkage was determined as follows:  $^1\text{H}$  NMR signals of the C-8 methoxy group ( $\delta$  4.01) and H-10 ( $\delta$  7.71) were shifted downfield compared with those of **11** ( $\delta$  3.89, 7.30), suggesting that the sugar moiety was linked to C-9. This assignment was supported by the results of COLOC and NOESY experiments. The optical rotation was levorotatory ( $[\alpha]^{27}_D -74.5^\circ$ ), indicating that the absolute configuration of C-6a is *R* ( $\beta$ -H).<sup>20</sup> Therefore, **1** was identified as stesakine-9-*O*- $\beta$ -D-glucopyranoside.

Glycoside **2** was isolated as an amorphous powder, and the molecular formula was assigned as  $\text{C}_{24}\text{H}_{29}\text{NO}_7$  by FABMS. In the EIMS, the molecular ion peak at  $m/z$  443 and strong ion peaks at  $m/z$  281  $[\text{M} - 162]^+$  and 280  $[\text{M} - 163]^+$  suggested that **2** should be also an alkaloid glycoside.<sup>17,18</sup> The  $^1\text{H}$  NMR spectrum exhibited signals of one *N*-methyl group ( $\delta$  2.41), one methoxy group ( $\delta$  3.93), four overlapping aromatic protons ( $\delta$  7.30–7.44), one separate aromatic proton at  $\delta$  8.70, and signals assigned to the sugar moiety. The  $^{13}\text{C}$  NMR spectrum showed 24 signals, including six signals assignable to  $\beta$ -glucopyranose. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra, except for signals of the sugar

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moiety, were closely related to those of *N*-methylasimilobine (**12**), indicating that **2** was a  $\beta$ -glucopyranoside of **12**. The sugar moiety was placed at C-2, because  $^1\text{H}$  NMR signals of the C-1 methoxy group ( $\delta$  3.93) and H-3 ( $\delta$  7.39) were shifted downfield compared with those of **12** ( $\delta$  3.73, 7.01). The HMBC and NOESY experiments also supported placement of the sugar moiety at C-2. The absolute configuration was C-6a*R* ( $\beta$ -H) as deduced from the optical rotation ( $[\alpha]_{\text{D}}^{27} - 116.1^\circ$ ).<sup>20</sup> Thus, **2** was assigned as *N*-methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside.

The structures of known alkaloids **3–14** were confirmed by comparison of the spectroscopic data with published values and from the results of 2D NMR experiments; the other known alkaloids were identified by direct comparison of the spectroscopic data with those of authentic samples isolated previously.<sup>1</sup> The aporphine alkaloids, including a dehydroaporphine and aporphine glycoside, were major constituents in the seeds of *S. cepharantha*. However, aporphines **1–13** were not detectable in the root tubers, leaves, and stems.

To date, only four aporphine glycosides have been reported: (–)-asimilobine-2-*O*- $\beta$ -D-glucoside,<sup>18</sup> kamaline,<sup>19</sup> floripavidine,<sup>21</sup> and tuberosinone-*N*- $\beta$ -D-glucoside.<sup>22</sup> The aglycons of these glycosides were asimilobine derivatives and tuberosinone, respectively. Therefore, stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**) represents the first aporphine glycoside having stesakine (**11**) as the aglycon. *N*-Methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**) was a fourth asimilobine derivative glycoside.

## Experimental Section

**General Experimental Procedures.** Melting points were determined on a Yanagimoto hot-stage melting point apparatus without correction. Optical rotations ( $[\alpha]_{\text{D}}$ ) were determined on a DIP-1000 (JASCO) polarimeter. IR spectra were obtained on an FT/IR-5000 (JASCO) spectrometer using KBr disks. NMR spectra were recorded on a JNM- $\alpha$ 500 (JEOL) (500 MHz for  $^1\text{H}$  and 125 MHz for  $^{13}\text{C}$ ) spectrometer with tetramethylsilane as internal standard. MS were recorded on a JMS-HX110A (JEOL) spectrometer for EIMS and FABMS, and a JMS-D300 (JEOL) spectrometer was used for HRMS. EIMS were measured at 60 eV. Column chromatography was performed on Wakogel C-200 (Wako Pure Chemical Industries, Ltd.). Preparative TLC was performed on precoated Si gel 60 F<sub>254</sub> (0.25 mm thickness) plates (Merck).

**Plant Material.** *S. cepharantha* was cultivated at Yasato-machi, Ibaraki Prefecture, in Japan, and the seeds were collected in October 1980. A voucher specimen was deposited at the research laboratory of Kaken Shoyaku Co., Ltd.

**Extraction and Isolation.** Dried and crushed seeds of *S. cepharantha* (1.07 kg) were extracted exhaustively with hot *n*-hexane (15 L), then with hot MeOH (15 L). After concentration in vacuo, the residue from *n*-hexane was 252 g, and the MeOH residue was 60 g. The *n*-hexane residue was subjected to column chromatography using *n*-hexane–benzene (1:1), benzene, and MeOH–CHCl<sub>3</sub> (1:1) as eluents, to afford fractions of 165 g, 45 g, and 42 g, respectively. The 45-g fraction was subjected to a combination of column chromatography and crystallization to afford dehydrostephanine (**3**, 1.1 g).<sup>4</sup> The 42-g fraction was subjected to a combination of column chromatography, preparative TLC, and crystallization, to give dehydrocrebanine (**4**, 28 mg),<sup>6</sup> stephanine (**5**, 382 mg),<sup>7</sup> crebanine (**6**, 535 mg),<sup>7</sup> (–)-dicentrine (7, 56 mg),<sup>8</sup> (–)-roemerine (**8**, 6 mg),<sup>9</sup> (–)-isolaureline (**9**, 10 mg),<sup>10</sup> nuciferine (**10**, 2 mg),<sup>11</sup> stesakine (**11**, 14 mg),<sup>6</sup> and *N*-methylasimilobine (**12**, 42 mg).<sup>11</sup> The MeOH residue was dissolved in 5% HCl. After filtration of insoluble materials, the solution was washed with Et<sub>2</sub>O. The aqueous layer was then adjusted to pH 10 with NH<sub>4</sub>OH and extracted successively with Et<sub>2</sub>O and CHCl<sub>3</sub>, to afford fractions **A** (142 mg) and **B** (144 mg), respectively. Fraction **A** was

subjected to column chromatography using 1%, 5%, and 20% MeOH–CHCl<sub>3</sub> as eluents to give fractions of 55 mg, 51 mg, and 38 mg, respectively. The first fraction (55 mg) was separated by preparative TLC to yield **6** (8 mg), **11** (18 mg), cepharanthine (4 mg),<sup>14</sup> and isotetrandrine (5 mg).<sup>16</sup> The 51-mg fraction was also purified by preparative TLC to afford **12** (18 mg), (–)-cassythicine (**13**, 4 mg),<sup>12</sup> pronuciferine (**14**, 2 mg),<sup>13</sup> cepharanoline (3 mg),<sup>15</sup> and berbamine (12 mg).<sup>17</sup> The final fraction (38 mg) was separated by preparative TLC [with EtOAc–MeOH (1:2)] to afford stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**, 5 mg) and *N*-methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**, 10 mg). Fraction **B** was subjected to column chromatography using 3%, 20%, and 50% MeOH–CHCl<sub>3</sub> and final purification by preparative TLC [with EtOAc–MeOH (1:2)] to afford **1** (25 mg) and **2** (30 mg).

The structures of known alkaloids **3–14** were confirmed by comparison of the spectroscopic data with published values and from the results of 2D NMR experiments; the other known alkaloids were identified by direct comparison of the spectroscopic data with those of authentic samples isolated previously.<sup>1</sup>

**Stesakine-9-*O*- $\beta$ -D-glucopyranoside (**1**):** amorphous powder;  $[\alpha]_{\text{D}}^{27} - 74.5^\circ$  (*c* 0.20, MeOH); IR (KBr)  $\nu_{\text{max}}$  3400, 1604, 1577, 1493, 1408, 1389, 1236, 1076, 1036 cm<sup>-1</sup>;  $^1\text{H}$  NMR (pyridine-*d*<sub>5</sub>, 500 MHz)  $\delta$  6.60 (s, H-3), 2.51 (ddd, *J* = 16.2, 3.7, 1.2 Hz, H-4), 3.12 (ddd, *J* = 16.2, 10.7, 5.5 Hz, H-4), 2.39 (ddd, *J* = 11.6, 10.7, 3.7 Hz, H-5), 2.94 (ddd, *J* = 11.6, 5.5, 1.2 Hz, H-5), 3.08 (dd, *J* = 13.7, 4.3 Hz, H-6a), 2.48 (dd, *J* = 13.7, 13.7 Hz, H-7), 3.91 (dd, *J* = 13.7, 4.3 Hz, H-7), 7.71 (d, *J* = 8.6 Hz, H-10), 8.01 (d, *J* = 8.6 Hz, H-11), 5.98 (d, *J* = 1.2 Hz, OCH<sub>2</sub>O), 6.11 (d, *J* = 1.2 Hz, OCH<sub>2</sub>O), 2.48 (s, *N*-CH<sub>3</sub>), 4.01 (s, 8-OCH<sub>3</sub>), 5.71 (d, *J* = 7.3 Hz, H-1'), 4.38 (m, H-2'), 4.38 (m, H-3'), 4.35 (m, H-4'), 4.19 (ddd, *J* = 8.9, 5.5, 2.1 Hz, H-5'), 4.42 (dd, *J* = 11.9, 5.5 Hz, H-6'), 4.60 (dd, *J* = 11.9, 2.1 Hz, H-6');  $^{13}\text{C}$  NMR (pyridine-*d*<sub>5</sub>, 125 MHz)  $\delta$  142.7 (s, C-1), 147.12 (s, C-2), 107.4 (d, C-3), 127.32 (s, C-3a), 29.6 (t, C-4), 53.9 (t, C-5), 62.47 (d, C-6a), 27.5 (t, C-7), 130.3 (s, C-7a), 147.07 (s, C-8), 151.2 (s, C-9), 115.4 (d, C-10), 123.8 (d, C-11), 126.3 (s, C-11a), 116.9 (s, C-11b), 127.25 (s, C-11c), 101.1 (t, OCH<sub>2</sub>O), 44.0 (q, *N*-CH<sub>3</sub>), 61.0 (q, 8-OCH<sub>3</sub>), 102.7 (d, C-1'), 75.0 (t, C-2'), 78.8 (t, C-3'), 71.3 (t, C-4'), 79.1 (t, C-5'), 62.45 (t, C-6'); NOESY H-3–H-4; *N*-CH<sub>3</sub>–H-5, -6a, -7; H-7–8-OCH<sub>3</sub>; H-1'–H-10, -3', -5'; COLOC H-3 → C-1, -4, -11c; H-5 → C-3a, -6a; H-7 → C-8, -11a, -11c; H-10 → C-8, -11a; H-11 → C-7a, -9; OCH<sub>2</sub>O → C-1, -2, *N*-CH<sub>3</sub> → C-5, -6a; 8-OCH<sub>3</sub> → C-8; H-1' → C-9; EIMS *m/z* 487 [M]<sup>+</sup> (7), 485 (8), 325 (64), 324 (100), 308 (19), 282 (34), 280 (10), 267 (6), 264 (7), 251(7); positive FABMS *m/z* 488 [M + H]<sup>+</sup>; positive HRFABMS *m/z* 488.1935 [M + H]<sup>+</sup> (calcd for C<sub>25</sub>H<sub>30</sub>NO<sub>9</sub>, 488.1920).

***N*-Methylasimilobine-2-*O*- $\beta$ -D-glucopyranoside (**2**):** amorphous powder;  $[\alpha]_{\text{D}}^{27} - 116.1^\circ$  (*c* 0.60, MeOH); IR (KBr)  $\nu_{\text{max}}$  3400, 1595, 1573, 1496, 1425, 1371, 1254, 1074, 1045 cm<sup>-1</sup>;  $^1\text{H}$  NMR (pyridine-*d*<sub>5</sub>, 500 MHz)  $\delta$  7.39 (s, H-3), 2.50 (dd, *J* = 17.7, 3.4 Hz, H-4), 3.11 (ddd, *J* = 17.7, 11.9, 5.5 Hz, H-4), 2.23 (ddd, *J* = 11.9, 11.6, 3.4 Hz, H-5), 2.84 (dd, *J* = 11.6, 5.5 Hz, H-5), 2.96 (dd, *J* = 13.7, 4.0 Hz, H-6a), 2.75 (dd, *J* = 13.7, 13.7 Hz, H-7), 3.21 (dd, *J* = 13.7, 4.0 Hz, H-7), 7.38 (d, *J* = 7.6 Hz, H-8), 7.31 (dd, *J* = 7.6, 7.6 Hz, H-9), 7.42 (dd, *J* = 7.9, 7.6 Hz, H-10), 8.70 (d, *J* = 7.9 Hz, H-11), 2.41 (s, *N*-CH<sub>3</sub>), 3.93 (s, 1-OCH<sub>3</sub>), 5.64 (d, *J* = 7.0 Hz, H-1'), 4.38 (m, H-2'), 4.37 (m, H-3'), 4.31 (m, H-4'), 4.16 (ddd, *J* = 7.6, 5.5, 2.1 Hz, H-5'), 4.40 (dd, *J* = 11.9, 5.5 Hz, H-6'), 4.62 (dd, *J* = 11.9, 2.1 Hz, H-6');  $^{13}\text{C}$  NMR (pyridine-*d*<sub>5</sub>, 125 MHz)  $\delta$  146.1 (s, C-1), 151.2 (s, C-2), 116.1 (d, C-3), 129.6 (s, C-3a), 29.6 (t, C-4), 53.5 (t, C-5), 62.9 (d, C-6a), 35.4 (t, C-7), 137.3 (s, C-7a), 128.6 (s, C-8), 127.8 (s, C-9), 127.3 (d, C-10), 128.8 (d, C-11), 132.9 (s, C-11a), 127.2 (s, C-11b), 129.9 (s, C-11c), 44.0 (q, *N*-CH<sub>3</sub>), 60.7 (q, 1-OCH<sub>3</sub>), 102.4 (d, C-1'), 75.0 (t, C-2'), 78.9 (t, C-3'), 71.4 (t, C-4'), 79.1 (t, C-5'), 62.6 (t, C-6'); NOESY H-3–H-4; *N*-CH<sub>3</sub>–H-5, -6a, -7; H-7–H-8, H-10–H-11; H-11–1-OCH<sub>3</sub>; H-1'–H-3, -3', -5'; HMBC H-3 → C-1, -4, -11c; H-5 → C-3a, -6a; H-7 → C-11a, -11c; H-8 → C-7, C-10, -11a; H-9 → C-7a, -11; H-10 → C-8, -11a; H-11 → C-7a, -9; 1-OCH<sub>3</sub> → C-1; *N*-CH<sub>3</sub> → C-5, -6a; H-1' → C-2, -5'; EIMS *m/z* 443 [M]<sup>+</sup> (36), 281 (73), 280 (100), 266 (50), 250 (31), 249 (29), 238 (25), 178 (18), 165 (20); positive FABMS *m/z* 444

[M + H]<sup>+</sup>; positive HRFABMS *m/z* 444.2010 [M + H]<sup>+</sup> (calcd for C<sub>24</sub>H<sub>30</sub>NO<sub>7</sub>, 444.2022).

**Stesakine (11):** mp 160–161° (MeOH); [α]<sub>D</sub><sup>27</sup> –89.8° (*c* 0.60, CHCl<sub>3</sub>); IR (KBr) *ν*<sub>max</sub> 3400, 1604, 1583, 1496, 1388, 1296, 1234, 1039 cm<sup>-1</sup>; <sup>1</sup>H NMR (pyridine-*d*<sub>5</sub>, 500 MHz) δ 6.60 (s, H-3), 2.52 (m, H-4), 3.12 (m, H-4), 2.40 (ddd, *J* = 12.1, 11.9, 4.0 Hz, H-5), 2.94 (ddd, *J* = 11.9, 5.8, 1.2 Hz, H-5), 3.11 (dd, *J* = 13.7, 4.3 Hz, H-6a), 2.54 (dd, *J* = 14.3, 13.7 Hz, H-7), 3.91 (dd, *J* = 13.4, 4.3 Hz, H-7), 7.30 (d, *J* = 8.6 Hz, H-10), 8.07 (d, *J* = 8.6 Hz, H-11), 6.00 (d, *J* = 1.2 Hz, OCH<sub>2</sub>O), 6.11 (d, *J* = 1.2 Hz, OCH<sub>2</sub>O), 2.50 (s, *N*-CH<sub>3</sub>), 3.89 (s, 8-OCH<sub>3</sub>), 11.69 (br s, 9-OH); <sup>13</sup>C NMR (pyridine-*d*<sub>5</sub>, 125 MHz) δ 142.4 (s, C-1), 147.1 (s, C-2), 106.9 (d, C-3), 127.3 (s, C-3a), 29.7 (t, C-4), 53.9 (t, C-5), 62.6 (d, C-6a), 27.8 (t, C-7), 130.4 (s, C-7a), 145.8 (s, C-8), 151.3 (s, C-9), 115.8 (d, C-10), 124.40 (d, C-11), 124.40 (s, C-11a), 117.6 (s, C-11b), 127.6 (s, C-11c), 101.0 (t, OCH<sub>2</sub>O), 44.1 (q, *N*-CH<sub>3</sub>), 60.5 (q, 8-OCH<sub>3</sub>); EIMS *m/z* 325 [M]<sup>+</sup> (67), 324 (100), 308 (68), 282 (30), 280 (34), 267 (12), 264 (17), 251 (15), 222 (14); HRMS *m/z* 325.1313 (calcd for C<sub>19</sub>H<sub>19</sub>NO<sub>4</sub>, 325.1313).

***N*-Methylasimilobine (12):** mp 195–197° (EtOAc); [α]<sub>D</sub><sup>27</sup> –211.2° (*c* 0.66, CHCl<sub>3</sub>); IR 3400, 1610, 1589, 1570, 1496, 1471, 1452, 1423, 1373, 1298, 1273, 1242, 1173, 1142 cm<sup>-1</sup>; <sup>1</sup>H NMR (pyridine-*d*<sub>5</sub>, 500 MHz) δ 7.01 (s, H-3), 2.57 (dd, *J* = 17.1, 3.4 Hz, H-4), 3.14 (ddd, *J* = 17.1, 11.9, 4.9 Hz, H-4), 2.36 (ddd, *J* = 11.9, 11.6, 3.4 Hz, H-5), 2.90 (dd, *J* = 11.6, 4.9 Hz, H-5), 3.01 (dd, *J* = 13.7, 3.7 Hz, H-6a), 2.78 (dd, *J* = 13.7, 13.7 Hz, H-7), 3.22 (dd, *J* = 13.7, 3.7 Hz, H-7), 7.40 (d, *J* = 7.3 Hz, H-8), 7.32 (dd, *J* = 7.6, 7.3 Hz, H-9), 7.43 (dd, *J* = 8.2, 7.6 Hz, H-10), 8.72 (d, *J* = 8.2 Hz, H-11), 2.43 (s, *N*-CH<sub>3</sub>), 3.73 (s, 1-OCH<sub>3</sub>), 11.18 (br s, 2-OH); <sup>13</sup>C NMR (pyridine-*d*<sub>5</sub>, 75 MHz) δ 145.0 (s, C-1), 150.9 (s, C-2), 116.6 (d, C-3), 130.1 (s, C-3a), 29.5 (t, C-4), 53.7 (t, C-5), 63.0 (d, C-6a), 35.6 (t, C-7), 137.3 (s, C-7a), 128.7 (d, C-8), 127.7 (d, C-9), 127.4 (d, C-10), 128.3 (d, C-11), 133.2 (s, C-11a), 126.9 (s, C-11b), 127.5 (s, C-11c), 44.1 (q, *N*-CH<sub>3</sub>), 60.00 (q, 1-OCH<sub>3</sub>); EIMS *m/z* 281 [M]<sup>+</sup> (100), 280 (69), 266 (37), 265 (37), 264 (29), 250 (39), 248 (22), 238 (19), 223 (14), 220 (14), 178 (29), 165 (19); HRMS *m/z* 281.1398 (calcd for C<sub>18</sub>H<sub>19</sub>NO<sub>2</sub>, 281.1413).

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## References and Notes

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